

# H1foo Has a Pivotal Role in Qualifying Induced Pluripotent Stem Cells

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## H1foo Has a Pivotal Role in Qualifying Induced Pluripotent Stem Cells

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### SUMMARY

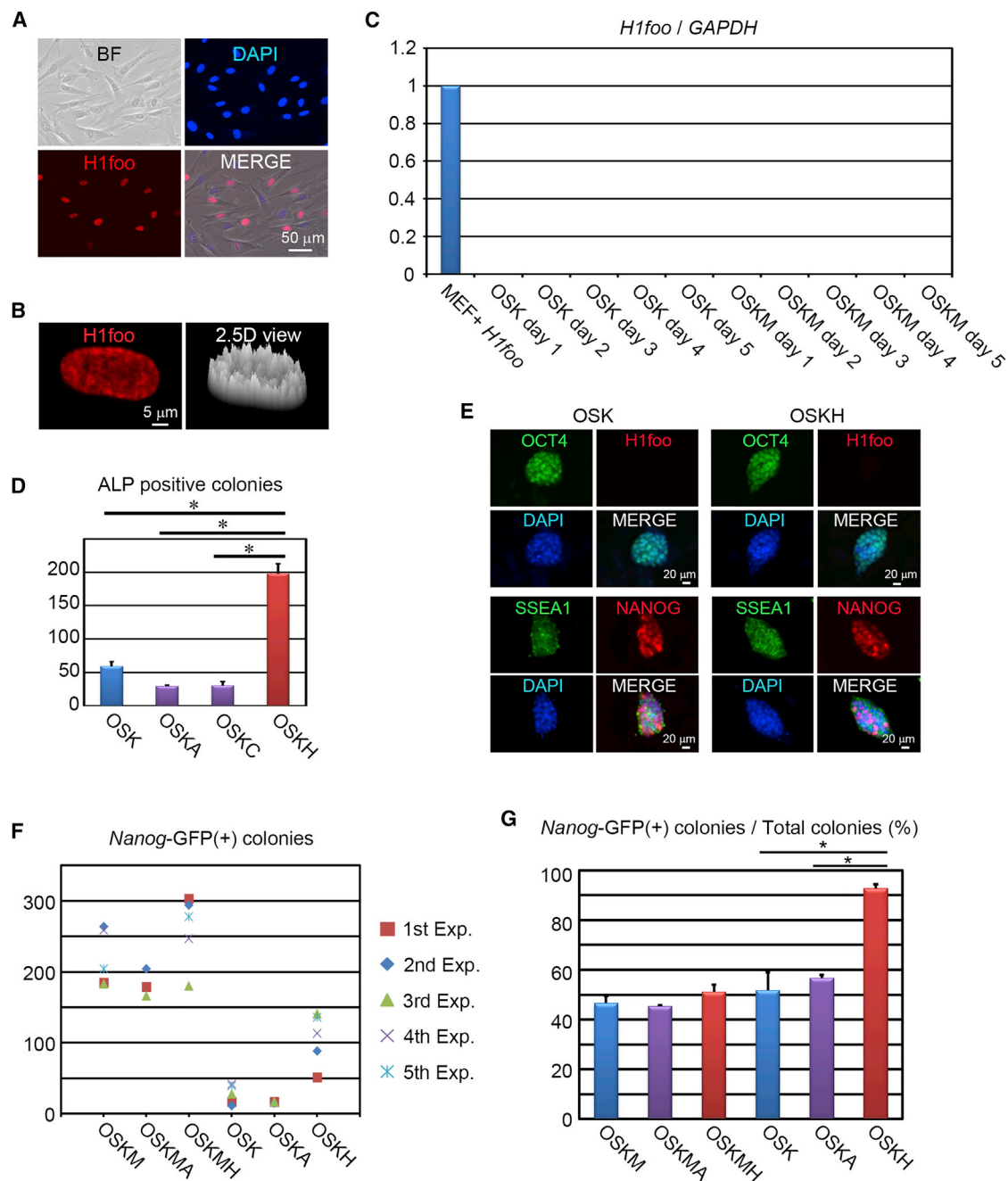
Embryonic stem cells (ESCs) are a hallmark of ideal pluripotent stem cells. Epigenetic reprogramming of induced pluripotent stem cells (iPSCs) has not been fully accomplished. iPSC generation is similar to somatic cell nuclear transfer (SCNT) in oocytes, and this procedure can be used to generate ESCs (SCNT-ESCs), which suggests the contribution of oocyte-specific constituents. Here, we show that the mammalian oocyte-specific linker histone *H1foo* has beneficial effects on iPSC generation. Induction of *H1foo* with *Oct4*, *Sox2*, and *Klf4* significantly enhanced the efficiency of iPSC generation. *H1foo* promoted in vitro differentiation characteristics with low heterogeneity in iPSCs. *H1foo* enhanced the generation of germline-competent chimeric mice from iPSCs in a manner similar to that for ESCs. These findings indicate that *H1foo* contributes to the generation of higher-quality iPSCs.

### INTRODUCTION

Induced pluripotent stem cells (iPSCs) can be generated from somatic cells by introducing *Oct4*, *Sox2*, *Klf4*, and *c-Myc* (Takahashi and Yamanaka, 2006). This can be achieved by reprogramming the transcription network and epigenetic signature of the parental somatic cells. iPSCs have several benefits for basic research, drug innovation, and regeneration therapy. However, recent studies have reported genetic and epigenetic variations with iPSCs (Hussein et al., 2011; Kim et al., 2010; Polo et al., 2010; Ruiz et al., 2012; Stadtfeld et al., 2010; Taapken et al., 2011), which influences gene expression and could lead to functional diversity within iPSC replicates (Liang and Zhang, 2013). In fact, several studies have reported the heterogeneous differentiation potential among generated iPSC clones compared with those of embryonic stem cells (ESCs) (Feng et al., 2010; Hu et al., 2010; Narsinh et al., 2011). It is important that every iPSC clone shows high quality without variation for basic research and clinical purposes. Many attempts have been made to solve these problems by various methods (Gafni et al., 2013), but not all iPSCs exhibit quality as high as that of ESCs.

The reprogramming of somatic cells was originally demonstrated by producing cloned frogs using somatic cell nuclear transfer (SCNT) into *Xenopus* oocytes (Gurdon

et al., 1958). Reprogramming of mammalian somatic cells has also been achieved using SCNT into oocytes, including those of mice and humans (Noggle et al., 2011; Wakayama et al., 1998). The procedure of SCNT into oocytes is similar to iPSC generation with respect to the time course of extinction of the parental gene-expression profile and activation of pluripotency (Egli et al., 2011). Both somatic cell reprogramming processes involve dynamic rearrangement of the epigenetic profile (Apostolou and Hochedlinger, 2013; Hussein et al., 2014). These findings suggest that the constituents of oocytes include a reprogramming-promoting factor. The linker histone H1 family binds to linker DNA and generates higher-order chromatin structures to control gene expression. Most members of the linker histone family consist of somatic linker histones that condense chromatin; therefore, these structures generally repress global gene-transcription activity (Hebbard and Archer, 2008; Steinbach et al., 1997). Mammalian oocytes contain the maternal-specific linker histone *H1foo*, a homolog of the *Xenopus* linker histone *B4*. *H1foo* is specifically expressed during the germinal vesicle stage and until the late two-cell or early four-cell stage, coincident with the early wave of zygotic genome reactivation (Gao et al., 2004; Tanaka et al., 2003, 2005). In *Xenopus* SCNT, somatic linker histones in transplanted nuclei are rapidly exchanged for linker histone *B4*, and the transplanted



**Figure 1. Exogenous Expression of *H1foo* Promotes iPSC Generation**

(A) Immunostaining for *H1foo* (red), including nuclei stained with DAPI (blue) and bright-field (BF) images. Scale bar, 50  $\mu$ m.

(B) Immunostaining for *H1foo* (red) with a 2D image (left) and a 2.5D image (right). The density of *H1foo* is depicted by the height in the 2.5D view. Scale bar, 5  $\mu$ m.

(C) qRT-PCR analysis of endogenous *H1foo* expression during reprogramming, normalized by *H1foo* expression in *H1foo*-overexpressed MEFs ( $n = 3$  independent experiments).

(D) Alkaline phosphatase-positive iPSC colony formations for OSK, OSK with *H1a* (OSK + *H1a*; OSKA), OSK with *H1c* (OSK + *H1c*; OSKC), and OSK with *H1foo* (OSK + *H1foo*; OSKH). Error bars represent the SEM ( $n = 3$  independent experiments). \* $p < 0.05$ .

(legend continued on next page)



nuclei begin to swell and initiate decondensation (Byrne et al., 2003; Jullien et al., 2010); furthermore, in mouse SCNT the same phenomenon is observed with *H1foo* (Becker et al., 2005). Unlike other somatic linker histones, *B4* and *H1foo* do not restrict the accessibility of the linker DNA, but decondense the chromatin and permit transcriptional activation (Hayakawa et al., 2012; Saeki et al., 2005).

Based on these studies, we hypothesized that *H1foo* has a beneficial effect on iPSC generation. Here, we show that *H1foo* enhanced the generation of mouse iPSCs when co-expressed with *Oct4*, *Sox2*, and *Klf4*. Furthermore, *H1foo* promoted several in vitro differentiation characteristics with low heterogeneity in iPSCs that were similar to those of ESCs. Specifically, *H1foo* enhanced germline-competent chimeric mouse generation. These findings indicate that *H1foo* contributes to the generation of higher-quality iPSCs.

## RESULTS

### Exogenous Expression of *H1foo* Promotes Qualified iPSC Generation

We examined the effect of exogenous *H1foo* on somatic cell reprogramming by introducing *H1foo* during iPSC generation with *Oct4*, *Sox2*, and *Klf4* (OSK) or *Oct4*, *Sox2*, *Klf4*, and *c-Myc* (OSKM). Retrovirus vector-mediated exogenous *H1foo* was exclusively expressed in the nucleus of adult mouse tail-tip fibroblasts (Figures 1A and S1A) and highly expressed peripherally in the nucleus (Figure 1B). SCNT into oocytes induced swelling of nuclei and chromatin decondensation (Gao et al., 2004; Tamada et al., 2006; Teranishi et al., 2004), so we investigated the effect of *H1foo* on nuclei (Figure S1B). We found that *H1a*, *H1c*, and *H1foo* had no significant effect on nuclear swelling (Figures S1C and S1D). However, interestingly, only *H1foo* reduced the intensively stained area, namely the heterochromatin area (Figure S1E). Next, we addressed whether intrinsic *H1foo* might be expressed during iPSC generation from mouse embryonic fibroblasts (MEFs) by introducing OSK or OSKM. However, we did not observe detectable intrinsic *H1foo* expression (Figure 1C). Co-expression of *H1foo* with OSK (OSKH) significantly enhanced the number of alkaline phosphatase-positive ESC-like colonies compared with OSK, OSK and *H1a* (OSKA), or OSK and *H1c* (OSKC) (Figure 1D). The OSKH-iPSCs expressed pluripotency markers similarly to control iPSCs (OSK), and *H1foo* was silenced (Figure 1E). We then examined the effect of *H1foo* on qualified iPSC generation

using tail-tip fibroblasts from *Nanog*-GFP transgenic adult mice (Okita et al., 2007) (Figure S1G). Interestingly, *H1foo* also maximally promoted *Nanog*-GFP-positive colony generation (8-fold) during iPSC generation by OSK (Figure 1F). Notably, *H1foo* specifically enhanced GFP-positive colonies as opposed to GFP-negative colonies (Figure 1G).

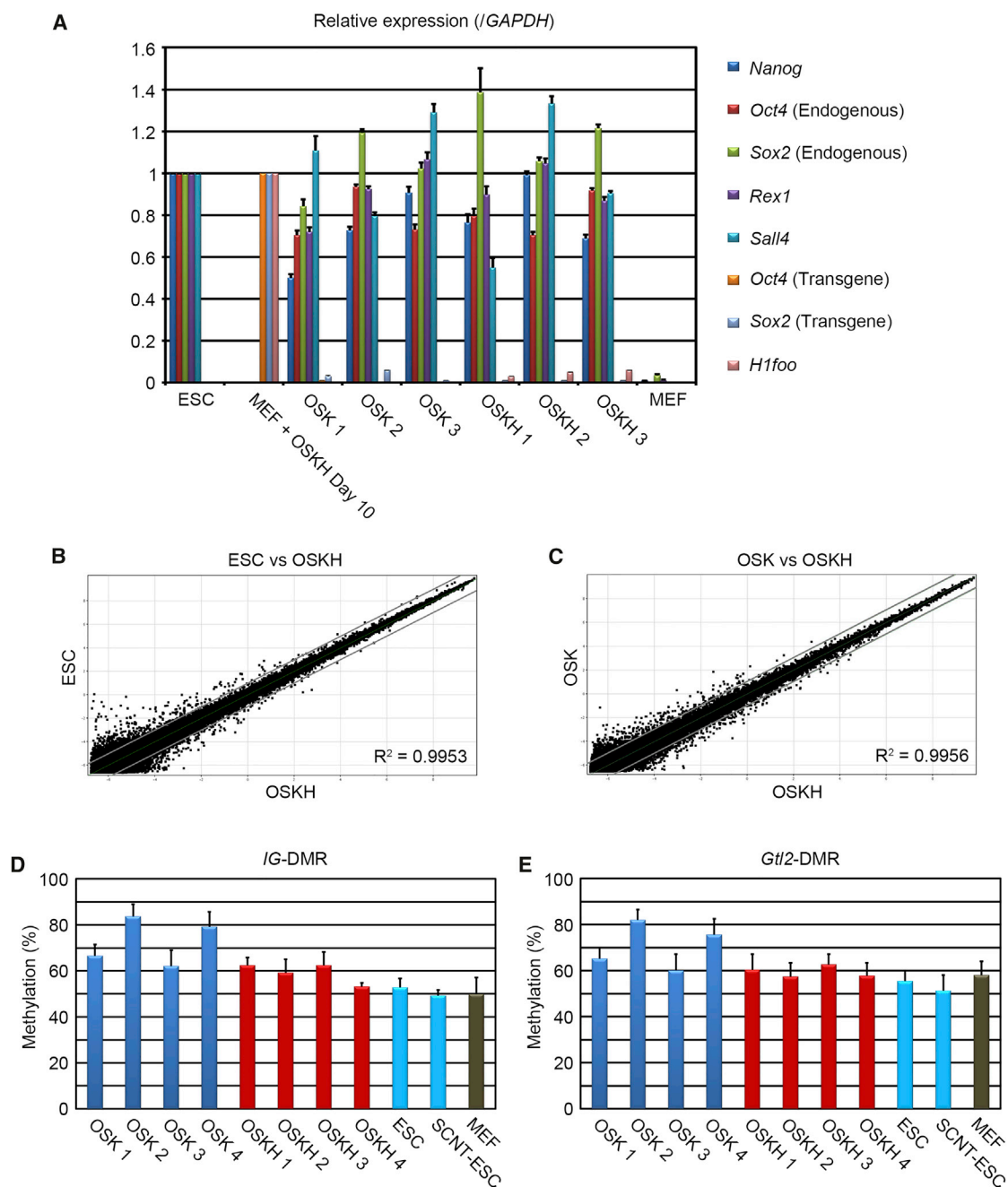
### Characteristics of OSKH-iPSC Generation

Next, we examined the iPSC characteristics produced by OSK and OSKH. The OSKH-iPSCs expressed pluripotency markers similar to those of OSK-iPSCs. Meanwhile the transgenes, including *H1foo*, were silenced (Figure 2A). Regarding the growth rate of iPSCs, there was no significant difference between OSK- and OSKH-treated cells (Figure S1F). We investigated the differences in global gene-transcriptome profiles among ESCs, three replicates of OSK-iPSCs, and four replicates of OSKH-iPSCs. All cell types were remarkably similar and showed a correlation coefficient ( $R^2$ ) of 0.99 (Figures 2B and 2C). We then examined DNA demethylation in the promoter regions of pluripotency marker genes (Figure S1H) and the differentiation potencies by teratoma formation (Figure S1I). Furthermore, we focused on genes differentially expressed between OSK-iPSCs and OSKH-iPSCs. We chose eight differentially expressed genes, which were statistically significant with more than a 2-fold difference between OSK-iPSCs and OSKH-iPSCs ( $p < 0.05$ ). Many of the genes showed more similar expression patterns between OSKH-iPSCs and ESCs than between OSK-iPSCs and ESCs (Figures S2A–S2H), but the expression levels of these transcripts were heterogeneous among the replicates in the same cell type, indicating that those genes would not have a definitive role in stem cell properties. Although a definitive marker that reflects the quality of iPSCs has not been found, aberrant epigenetic silencing of the *Dlk1-Dio3* gene cluster could indicate developmental potency, particularly because it contributes to the ratio of chimerism in mouse iPSCs (Stadtfield et al., 2010). We analyzed the methylation status of an intergenic differentially methylated region (IG-DMR) that is located between the *Dlk1* and *Gtl2* genes and a *Gtl2* differentially methylated region (*Gtl2*-DMR) (Figures 2D and 2E). Both loci in OSKH-iPSCs were highly demethylated, similarly to ESCs and MEFs. We next investigated the expression of several transcripts that increase or diminish in the early reprogramming phase (Lujan et al., 2015). Interestingly, OSKH-induced MEFs showed significant upregulation of several early reprogramming

(E) Immunostaining for pluripotency markers (OCT4 [green], SSEA1 [green], and Nanog [red]), including *H1foo* (red) and nuclei stained with DAPI (blue) in OSK-iPSCs and OSKH-iPSCs. Scale bars, 20  $\mu$ m.

(F) Number of *Nanog*-GFP-positive iPSC colonies in each experiment ( $n = 5$  independent experiments).

(G) Proportion of *Nanog*-GFP-positive colonies to the total number of ESC-like colonies. Error bars represent the SEM ( $n = 5$  independent experiments). \* $p < 0.05$ .



**Figure 2. Characteristics of OSKH-Induced iPSCs**

(A) qRT-PCR analysis of pluripotency genes and *H1foo* in iPSCs. Error bars represent the SEM ( $n = 3$  independent experiments). (B and C) Pairwise scatterplots of global gene-expression cDNA microarray patterns of OSKH-iPSCs ( $n = 4$ ) compared with ESC ( $n = 1$ ) (B) or OSK-iPSCs ( $n = 3$ ) (C). The gray lines indicate  $\log_2$  2-fold changes in gene-expression levels between the paired cell types. (D and E) Degree of DNA methylation at IG-DMR and Gtl2-DMR in four OSK iPSC clones, four OSKH-iPSC clones, and ESCs, as well as MEFs analyzed by pyrosequencing. Error bars represent the SEM ( $n = 3$  independent experiments).

markers (Figures S2I, S2J, and S2K) and downregulation of fibroblast markers (Figures S2L and S2M) in comparison with OSK- or OSKA-induced MEFs. We confirmed that OSKH correctly traces the same pathway already reported.

#### H1foo Enhances the In Vitro Differentiation Potential

Next, we asked whether *H1foo* could improve the quality of iPSCs. Often iPSCs highly express pluripotency markers but show poor differentiation efficiency in vitro, including





characteristics such as a low number of embryoid bodies (EBs) and frazzled EBs with poor differentiation potential. Moreover, this undesired trend is highly apparent in low-passage iPSCs and is correlated to the remnant source cell epigenetic memories (Kim et al., 2010; Polo et al., 2010). We examined the differentiation potency by EB formation in low-passage (P5) OSK-iPSCs, OSKH-iPSCs, and two types of ESCs as controls. First, we examined the number and the size of EBs at 5 days after differentiation (Figure S3A), which were more similar between OSKH-iPSCs and ESCs than between OSK-iPSCs and ESCs (Figures 3A and 3B). Furthermore, the variance of EB size was smaller in OSKH-iPSCs than in OSK-iPSCs (Figures 3C and 3D). OSKH-iPSCs expressed a higher content of proliferation markers than OSK-iPSCs (Figures 3E and 3F). However, a significant difference was not observed in the differentiation marker profiles among ESCs, OSK-iPSCs, and OSKH-iPSCs (Figure S3B). We also examined the population of apoptotic cells during differentiation. Apoptosis in OSKH-iPSCs was suppressed compared with OSK-iPSCs (Figures 3G and S3C). Taken together, the data suggest that *H1foo* causes iPSCs to be more adaptable to in vitro differentiation conditions and promotes the homogeneity of EBs.

### H1foo Enhances the In Vivo Differentiation Potential

We finally investigated the in vivo differentiation potential of OSK-iPSCs and OSKH-iPSCs using a co-culture aggregation method (Eakin and Hadjantonakis, 2006). First we examined the chromosome number, which showed no significant difference in chromosomal abnormality between OSK-iPSCs and OSKH-iPSCs (Figure 4A). Each iPSC clone was aggregated with 100 ICR mouse embryos at the eight-cell stage, and chimeric embryos were transferred into the uteri of pseudopregnant females. Notably, OSKH-iPSCs generated more live chimeras, with higher chimerism, than OSK-iPSCs (Figures 4B, 4C, S4B, and S4C). Moreover, some OSKH-iPSC clones generated more 100% chimeras than ESCs (Figure S4B). Germline transmission potential is one of the most stringent hallmarks of pluripotent stem cells. We examined the germline transmission from 100% chimeric mice by in vitro fertilization. OSKH-iPSCs generated many pups with colored coats, confirming the favorable germline transmission potential of OSKH-iPSCs (Figures 4E, S4D, and S4E). We did not find any phenotypic abnormalities in these pups.

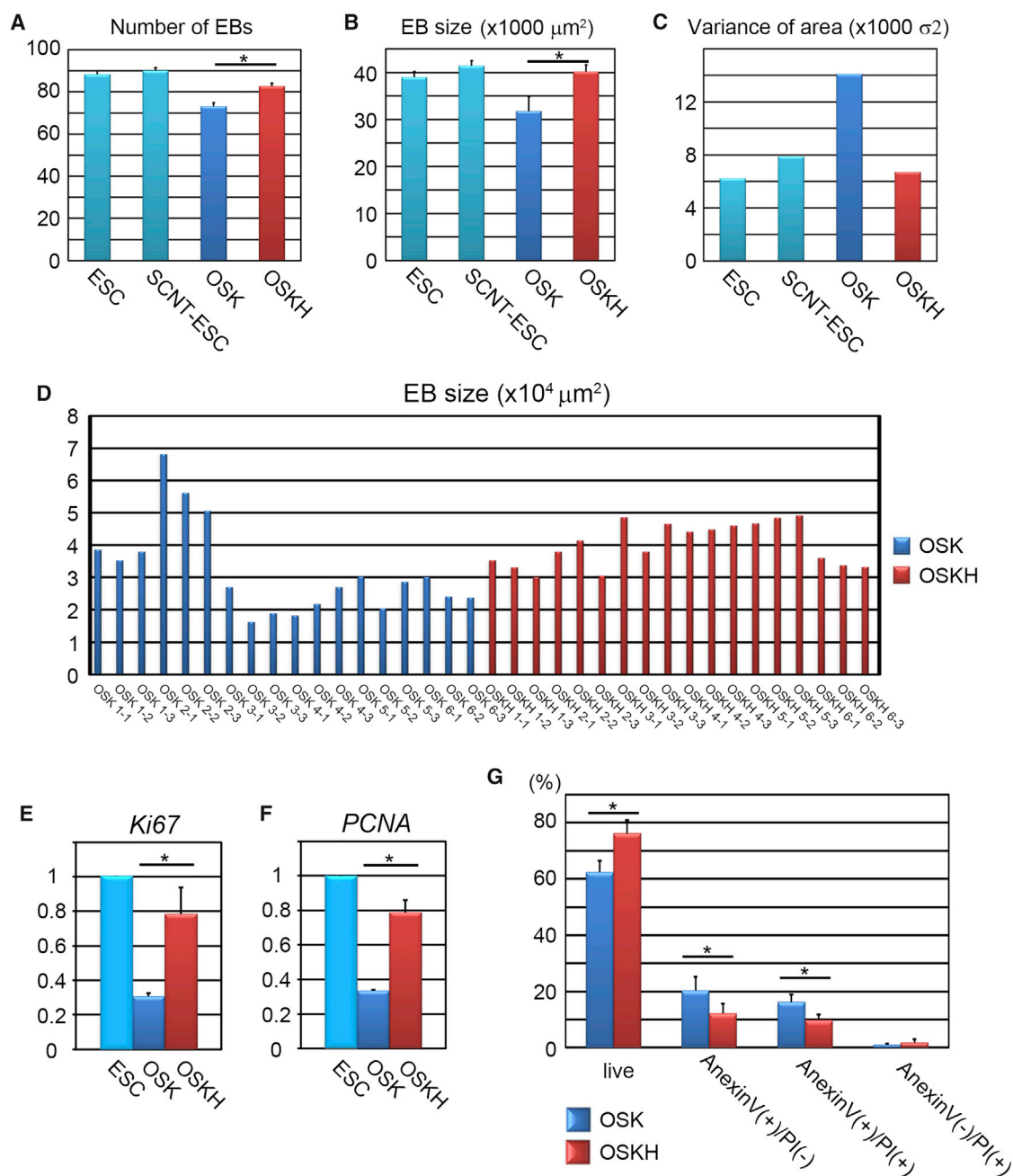
## DISCUSSION

*H1foo* promoted the generation of *Nanog*-GFP-positive colonies when it was co-expressed with OSK. Notably, the proportion of GFP-positive colonies was improved to 90%. OSKH-iPSCs demonstrated differentiation potency more

similar to that of ESCs than did OSK-iPSCs, especially with respect to in vitro EB formation, chimerism, and germline transmission in vivo. We also examined the effect of *H1foo* with *Oct4*, *Sox2*, *Klf4*, and *c-Myc*, but the number of alkaline phosphatase-positive colonies and *Nanog*-GFP colonies, and the proportion of GFP-positive colonies were not significantly different to those of OSKM-iPSCs. *c-Myc* promotes iPSC generation but is not essential for reprogramming. On the other hand, *c-Myc* lowers the proportion of *Nanog*-GFP-positive colonies and increases the tumorigenicity of cells (Nakagawa et al., 2008). Therefore, it is preferable to omit *c-Myc* in iPSC generation, although it is still commonly used to promote reprogramming efficiency. We showed here that *H1foo* could be substituted for *c-Myc* in terms of reprogramming efficiency and showed superiority with respect to qualifying iPSCs.

Recent studies have demonstrated that oocyte constituents play a key role in somatic cell reprogramming in SCNT. Co-expression of maternal-specific factors in oocytes, such as Glis1 (Maekawa et al., 2011) and TH2A/TH2B (Shinagawa et al., 2014), enhances the reprogramming efficiency of iPSC generation. Investigation of maternal-specific factors in oocytes has great potential for innovating somatic cell reprogramming and deciphering the reprogramming mechanisms (Gurdon and Wilmot, 2011). *H1foo* is specifically expressed during the germinal vesicle stage and is essential for oocyte maturation (Furuya et al., 2007; Gao et al., 2004). Interestingly, exogenous expression of *H1foo* in ESCs leads to the prevention of differentiation in vitro due to continuous pluripotency gene activation (Hayakawa et al., 2012). In our study, *H1foo* was properly silenced in generated iPSCs, which induced successful reprogramming but did not hinder the differentiation potency of OSKH-iPSCs.

The detailed molecular mechanisms regarding how *H1foo* enhances the reprogramming efficiency in iPSC generation and why OSKH-iPSCs exhibit improved quality remain elusive. The higher-order chromatin structure is crucially dependent on architectural chromatin proteins, including the family of linker histone proteins. Although somatic cells contain numerous linker histone variants, only one, *H1foo*, is present in mouse oocytes (Tanaka et al., 2001). In the mouse egg, somatic linker histones in sperm-derived chromatin are rapidly replaced by *H1foo* after fertilization (Tanaka et al., 2001). In SCNT oocytes, the somatic linker histone H1c in the donor chromatin is also rapidly replaced by *H1foo* in mice (Gao et al., 2004; Teranishi et al., 2004). In *Xenopus* SCNT, oocyte-specific linker histone B4 loading to genome-wide somatic chromatin is required for successful reprogramming (Jullien et al., 2010, 2014; Miyamoto et al., 2007). In the early phase of the reprogramming process, global loss of histone H3 lysine 27 trimethylation (H3K27me3) occurs and epigenetic modification affects



**Figure 3. In Vitro Differentiation Potential in OSKH-iPSCs**

(A) Number of EBs on day 5 after differentiation. Error bars represent the SEM (n = 3 independent experiments). \*p < 0.05.

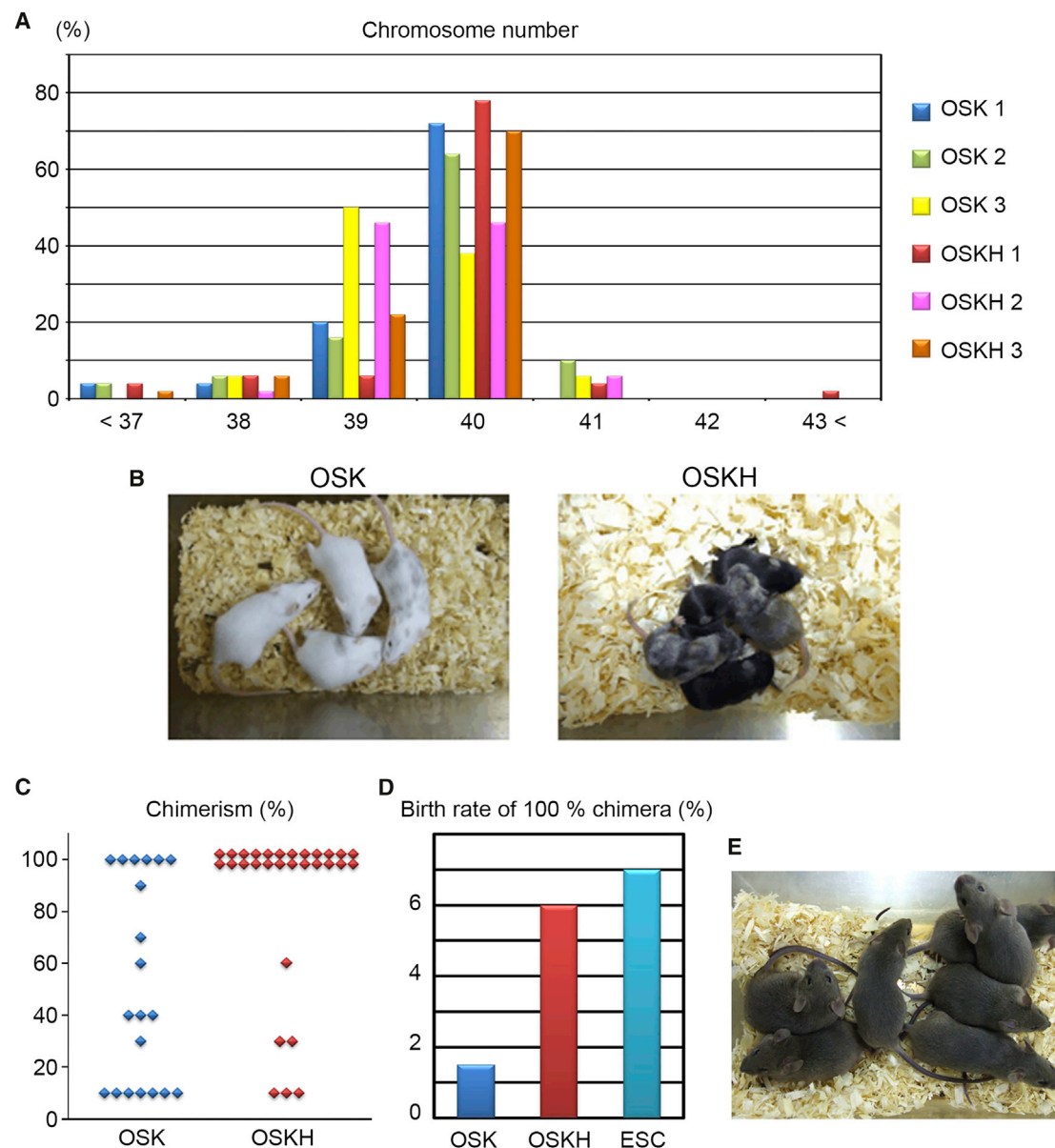
(B) Size of EBs on day 5 after differentiation. Error bars represent the SEM (n = 3 independent experiments). \*p < 0.05.

(C) Variance of EB sizes from each ESCs and iPSCs (n = 3 independent experiments).

(D) Variation of EB sizes. Each bar represents one experiment.

(E and F) Cell-proliferation markers Ki67 (E) and proliferating cell nuclear antigen (PCNA; F) on day 2 after differentiation. Error bars represent the SEM (n = 3 independent experiments). \*p < 0.05.

(G) Apoptotic cell distribution determined by fluorescence-activated cell sorting analysis of cells labeled with annexin V-fluorescein isothiocyanate and propidium iodide (PI) on day 1 after differentiation induction. Error bars represent the SEM (n = 3 independent experiments). \*p < 0.05.







*H1foo* induced successful reprogramming for iPSC generation in a stringent assay, thus contributing to chimerism and germline transmission. Although in vivo experiments cannot be performed in humans, it is important to generate high-quality iPSCs without variation among different iPSC lines.

## EXPERIMENTAL PROCEDURES

Details of all procedures are available in [Supplemental Experimental Procedures](#).

All experiments were performed in accordance with the Keio University Animal Care Guidelines and were approved by the Ethics Committee of Keio University (20-041-4), which conforms to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health.

## ACCESSION NUMBERS

The accession number for the microarray data reported in this paper is GEO: GSE79515.

## SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, four figures, and one table and can be found with this article online at <http://dx.doi.org/10.1016/j.stemcr.2016.04.015>.

## AUTHOR CONTRIBUTIONS

S.Y. designed the research. A.K., S.Y., F.S., Y.S., T.S., D.K., S.K., M.T., S.T., H.H., T.E., Y.T., S.M., S.T., H.O., K.Y., H.W., M.O., and R.K. performed the research. T.N. conducted the electron microscopic observation. A.K., S.Y., F.S., Y.M., H.O., K.Y., M.T., and K.F. analyzed the data. A.K. and S.Y. wrote the paper.

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## REFERENCES

- Apostolou, E., and Hochedlinger, K. (2013). Chromatin dynamics during cellular reprogramming. *Nature* 502, 462–471.
- Becker, M., Becker, A., Miyara, F., Han, Z., Kihara, M., Brown, D.T., Hager, G.L., Latham, K., Adashi, E.Y., and Misteli, T. (2005). Differential in vivo binding dynamics of somatic and oocyte-specific linker histones in oocytes and during ES cell nuclear transfer. *Mol. Biol. Cell* 16, 3887–3895.
- Byrne, J.A., Simonsson, S., Western, P.S., and Gurdon, J.B. (2003). Nuclei of adult mammalian somatic cells are directly reprogrammed to oct-4 stem cell gene expression by amphibian oocytes. *Curr. Biol.* 13, 1206–1213.
- Eakin, G.S., and Hadjantonakis, A.-K. (2006). Production of chimeras by aggregation of embryonic stem cells with diploid or tetraploid mouse embryos. *Nat. Protoc.* 1, 1145–1153.
- Egli, D., Chen, A.E., Saphier, G., Ichida, J., Fitzgerald, C., Go, K.J., Acevedo, N., Patel, J., Baetscher, M., Kearns, W.G., et al. (2011). Reprogramming within hours following nuclear transfer into mouse but not human zygotes. *Nat. Commun.* 2, 488.
- Feng, Q., Lu, S.J., Klimanskaya, I., Gomes, I., Kim, D., Chung, Y., Honig, G.R., Kim, K.S., and Lanza, R. (2010). Hemangioblastic derivatives from human induced pluripotent stem cells exhibit limited expansion and early senescence. *Stem Cells* 28, 704–712.
- Furuya, M., Tanaka, M., Teranishi, T., Matsumoto, K., Hosoi, Y., Saeki, K., Ishimoto, H., Minegishi, K., Iritani, A., and Yoshimura, Y. (2007). H1foo is indispensable for meiotic maturation of the mouse oocyte. *J. Reprod. Dev.* 53, 895–902.
- Gafni, O., Weinberger, L., Mansour, A.A., Manor, Y.S., Chomsky, E., Ben-Yosef, D., Kalma, Y., Viukov, S., Maza, I., Zviran, A., et al. (2013). Derivation of novel human ground state naive pluripotent stem cells. *Nature* 504, 282–286.
- Gao, S., Chung, Y.G., Parseghian, M.H., King, G.J., Adashi, E.Y., and Latham, K.E. (2004). Rapid H1 linker histone transitions following fertilization or somatic cell nuclear transfer: evidence for a uniform developmental program in mice. *Dev. Biol.* 266, 62–75.
- Gurdon, J.B., and Wilmut, I. (2011). Nuclear transfer to eggs and oocytes. *Cold Spring Harbor Perspect. Biol.* 3. <http://dx.doi.org/10.1101/cshperspect.a002659>.
- Gurdon, J.B., Elsdale, T.R., and Fischberg, M. (1958). Sexually mature individuals of *Xenopus laevis* from the transplantation of single somatic nuclei. *Nature* 182, 64–65.
- Hayakawa, K., Ohgane, J., Tanaka, S., Yagi, S., and Shiota, K. (2012). Oocyte-specific linker histone H1foo is an epigenomic modulator that decondenses chromatin and impairs pluripotency. *Epigenetics* 7, 1029–1036.
- Hebbar, P.B., and Archer, T.K. (2008). Altered histone H1 stoichiometry and an absence of nucleosome positioning on transfected DNA. *J. Biol. Chem.* 283, 4595–4601.
- Hu, B.Y., Weick, J.P., Yu, J., Ma, L.X., Zhang, X.Q., Thomson, J.A., and Zhang, S.C. (2010). Neural differentiation of human induced pluripotent stem cells follows developmental principles but with variable potency. *Proc. Natl. Acad. Sci. USA* 107, 4335–4340.
- Hussein, S.M., Batada, N.N., Vuoristo, S., Ching, R.W., Autio, R., Narva, E., Ng, S., Sourour, M., Hamalainen, R., Olsson, C., et al.



- (2011). Copy number variation and selection during reprogramming to pluripotency. *Nature* 471, 58–62.
- Hussein, S.M., Puri, M.C., Tonge, P.D., Benevento, M., Corso, A.J., Clancy, J.L., Mosbergen, R., Li, M., Lee, D.S., Cloonan, N., et al. (2014). Genome-wide characterization of the routes to pluripotency. *Nature* 516, 198–206.
- Jullien, J., Astrand, C., Halley-Stott, R.P., Garrett, N., and Gurdon, J.B. (2010). Characterization of somatic cell nuclear reprogramming by oocytes in which a linker histone is required for pluripotency gene reactivation. *Proc. Natl. Acad. Sci. USA* 107, 5483–5488.
- Jullien, J., Miyamoto, K., Pasque, V., Allen, G.E., Bradshaw, C.R., Garrett, N.J., Halley-Stott, R.P., Kimura, H., Ohsumi, K., and Gurdon, J.B. (2014). Hierarchical molecular events driven by oocyte-specific factors lead to rapid and extensive reprogramming. *Mol. Cell* 55, 524–536.
- Kim, K., Doi, A., Wen, B., Ng, K., Zhao, R., Cahan, P., Kim, J., Aryee, M.J., Ji, H., Ehrlich, L.I., et al. (2010). Epigenetic memory in induced pluripotent stem cells. *Nature* 467, 285–290.
- Liang, G., and Zhang, Y. (2013). Genetic and epigenetic variations in iPSCs: potential causes and implications for application. *Cell Stem Cell* 13, 149–159.
- Lujan, E., Zunder, E.R., Ng, Y.H., Goronzy, I.N., Nolan, G.P., and Wernig, M. (2015). Early reprogramming regulators identified by prospective isolation and mass cytometry. *Nature* 521, 352–356.
- Maekawa, M., Yamaguchi, K., Nakamura, T., Shibukawa, R., Kodanaka, I., Ichisaka, T., Kawamura, Y., Mochizuki, H., Goshima, N., and Yamanaka, S. (2011). Direct reprogramming of somatic cells is promoted by maternal transcription factor Glis1. *Nature* 474, 225–229.
- Miyamoto, K., Furusawa, T., Ohnuki, M., Goel, S., Tokunaga, T., Minami, N., Yamada, M., Ohsumi, K., and Imai, H. (2007). Reprogramming events of mammalian somatic cells induced by *Xenopus laevis* egg extracts. *Mol. Reprod. Dev.* 74, 1268–1277.
- Nakagawa, M., Koyanagi, M., Tanabe, K., Takahashi, K., Ichisaka, T., Aoi, T., Okita, K., Mochizuki, Y., Takizawa, N., and Yamanaka, S. (2008). Generation of induced pluripotent stem cells without Myc from mouse and human fibroblasts. *Nat. Biotechnol.* 26, 101–106.
- Narsinh, K.H., Sun, N., Sanchez-Freire, V., Lee, A.S., Almeida, P., Hu, S., Jan, T., Wilson, K.D., Leong, D., Rosenberg, J., et al. (2011). Single cell transcriptional profiling reveals heterogeneity of human induced pluripotent stem cells. *J. Clin. Invest.* 121, 1217–1221.
- Noggle, S., Fung, H.L., Gore, A., Martinez, H., Satriani, K.C., Prosser, R., Oum, K., Paull, D., Druckenmiller, S., Freeby, M., et al. (2011). Human oocytes reprogram somatic cells to a pluripotent state. *Nature* 478, 70–75.
- Okita, K., Ichisaka, T., and Yamanaka, S. (2007). Generation of germline-competent induced pluripotent stem cells. *Nature* 448, 313–317.
- Polo, J.M., Liu, S., Figueroa, M.E., Kulalert, W., Eminli, S., Tan, K.Y., Apostolou, E., Stadtfeld, M., Li, Y., Shioda, T., et al. (2010). Cell type of origin influences the molecular and functional properties of mouse induced pluripotent stem cells. *Nat. Biotechnol.* 28, 848–855.
- Ruiz, S., Diep, D., Gore, A., Panopoulos, A.D., Montserrat, N., Plongthongkum, N., Kumar, S., Fung, H.L., Giorgetti, A., Bilic, J., et al. (2012). Identification of a specific reprogramming-associated epigenetic signature in human induced pluripotent stem cells. *Proc. Natl. Acad. Sci. USA* 109, 16196–16201.
- Saeki, H., Ohsumi, K., Aihara, H., Ito, T., Hirose, S., Ura, K., and Kaneda, Y. (2005). Linker histone variants control chromatin dynamics during early embryogenesis. *Proc. Natl. Acad. Sci. USA* 102, 5697–5702.
- Shinagawa, T., Takagi, T., Tsukamoto, D., Tomaru, C., Huynh, L.M., Sivaraman, P., Kumarevel, T., Inoue, K., Nakato, R., Katou, Y., et al. (2014). Histone variants enriched in oocytes enhance reprogramming to induced pluripotent stem cells. *Cell Stem Cell* 14, 217–227.
- Stadtfeld, M., Apostolou, E., Akutsu, H., Fukuda, A., Follett, P., Natesan, S., Kono, T., Shioda, T., and Hochedlinger, K. (2010). Aberrant silencing of imprinted genes on chromosome 12qF1 in mouse induced pluripotent stem cells. *Nature* 465, 175–181.
- Steinbach, O.C., Wolffe, A.P., and Rupp, R.A. (1997). Somatic linker histones cause loss of mesodermal competence in *Xenopus*. *Nature* 389, 395–399.
- Taapken, S.M., Nisler, B.S., Newton, M.A., Sampsel-Barron, T.L., Leonhard, K.A., McIntire, E.M., and Montgomery, K.D. (2011). Karyotypic abnormalities in human induced pluripotent stem cells and embryonic stem cells. *Nat. Biotechnol.* 29, 313–314.
- Takahashi, K., and Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663–676.
- Tamada, H., Van Thuan, N., Reed, P., Nelson, D., Katoku-Kikyo, N., Wudel, J., Wakayama, T., and Kikyo, N. (2006). Chromatin decondensation and nuclear reprogramming by nucleoplasmin. *Mol. Cell Biol.* 26, 1259–1271.
- Tanaka, M., Hennebold, J.D., Macfarlane, J., and Adashi, E.Y. (2001). A mammalian oocyte-specific linker histone gene H1oo: homology with the genes for the oocyte-specific cleavage stage histone (cs-H1) of sea urchin and the B4/H1M histone of the frog. *Development* 128, 655–664.
- Tanaka, M., Kihara, M., Meczekalski, B., King, G.J., and Adashi, E.Y. (2003). H1oo: a pre-embryonic H1 linker histone in search of a function. *Mol. Cell Endocrinol.* 202, 5–9.
- Tanaka, M., Kihara, M., Hennebold, J.D., Eppig, J.J., Viveiros, M.M., Emery, B.R., Carrell, D.T., Kirkman, N.J., Meczekalski, B., Zhou, J., et al. (2005). H1FOO is coupled to the initiation of oocyte growth. *Biol. Reprod.* 72, 135–142.
- Teranishi, T., Tanaka, M., Kimoto, S., Ono, Y., Miyakoshi, K., Kono, T., and Yoshimura, Y. (2004). Rapid replacement of somatic linker histones with the oocyte-specific linker histone H1foo in nuclear transfer. *Dev. Biol.* 266, 76–86.
- Wakayama, T., Perry, A.C.F., Zuccotti, M., Johnson, K.R., and Yanagimachi, R. (1998). Full-term development of mice from enucleated oocytes injected with cumulus cell nuclei. *Nature* 394, 369–374.